

CASE REPORT

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# Changes in thrombosis-related parameters after AstraZeneca COVID-19 vaccination in a male volunteer: a case report

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## Abstract

**Background:** We speculated that subclinical thrombosis may occur frequently through crosstalk between immune/inflammatory reactions and hemostasis after corona virus disease-2019 (COVID-19) vaccination. To test this hypothesis, we measured thrombosis-related parameters after COVID-19 vaccination in a volunteer for 21 days.

**Case presentation:** The following parameters were measured in a 72-year-old Korean man at 1 day before vaccination and on days 1, 3, 7, 14, and 21 post vaccination (AstraZeneca COVID-19 vaccine: ChAdOx1-S/nCoV-19, CTMAV563): complete blood count, platelet indices, thrombin receptor-activating peptide-induced platelet aggregation, prothrombin time, activated partial thromboplastin time, D-dimer, thrombin-antithrombin III complex (TAT), plasmin- $\alpha$ 2 antiplasmin complex (PAP), von Willebrand factor (vWF) antigen and activity, plasminogen activator inhibitor-1 (PAI-1), protein C and protein S antigen and activity, lupus anticoagulant, fibrinogen degradation product, and plasminogen. We found that the TAT had significantly increased from 0.7 ng/mL (baseline) to 21.7 ng/mL (day 1). There was a transient increase in the PAI-1 level from 7.2 ng/mL (baseline) to 10.9 ng/mL (day 3), followed by a decrease in PAP level from 0.9 ng/mL (baseline) to 0.3  $\mu$ g/mL (day 7), suggesting that plasmin generation is suppressed by PAI-1.

**Conclusions:** Increased thrombotic factors (such as decreased protein S) and decreased fibrinolytic activity due to increased PAI-1 were potential factors causing thrombogenesis after COVID-19 vaccination. Sequential measurement of platelet indices, TAT, PAP, protein C, protein S, vWF, D-dimer, and PAI-1 following COVID-19 vaccination was informative.

**Keywords:** COVID-19 vaccine, Adverse drug reaction, Blood coagulation, Fibrinolysis, Case report

## Background

Coronavirus disease-2019 (COVID-19) vaccine-associated immune thrombosis and thrombocytopenia (VITT) have emerged as serious adverse effects of COVID-19 vaccines, particularly vector vaccines, such as those manufactured by Oxford/AstraZeneca and Janssen/Johnson & Johnson [1, 2]. However, it is uncertain whether VITT

is the only thrombosis-related disease that occurs after COVID-19 vaccination. We speculated that there may be many more unaccounted instances of subclinical thromboses, which may be triggered through crosstalk between hemostasis and inflammation/immune reactions via the release of various cytokines after COVID-19 vaccination [3].

To test our hypothesis, we measured thrombosis-related parameters sequentially for 21 days after COVID-19 vaccination in a male volunteer.

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## Case presentation

The volunteer was a 72-year-old Korean man (weight 62.5 kg; height 168 cm). Vital signs on the day before vaccination were: blood pressure, 120/80 mmHg; pulse rate, 70 beats/min; and body temperature, 36.8 °C. He was diagnosed with diabetes mellitus, hypertension, and hypercholesterolemia at the age of 45 years. His routine medications were atorvastatin (10 mg), aspirin (100 mg), diltiazem (180 mg), and long-acting insulin, which continued throughout the observation period. The baseline laboratory findings are presented in Table 1.

## Timeline

At 3 p.m. on 9 June 2021, the volunteer received the AstraZeneca COVID-19 vaccine (5 mL) in the left deltoid muscle (ChAdOx1-S/nCoV-19 [recombinant] vaccine, CTMAV563). Blood samples were obtained 1 day before vaccination (baseline) and on days 1, 3, 7, 14, and 21 post vaccination.

## Diagnostic assessment

The following parameters were measured in the context of routine testing in a selected general hospital: platelet indices (PI), including mean platelet volume (MPV) and platelet distribution width (PDW), and platelet-large cell ratio (P-LCR) were measured together on a fully automated hematology analyzer (Sysmex XN-1000<sup>TM</sup>; Sysmex Corp., Kobe, Japan). Thrombin receptor-activating peptide (TRAP)-induced platelet aggregation was measured using a Multiplate<sup>®</sup> analyzer (Roche Diagnostics GmbH, Mannheim, Germany). Prothrombin time, activated partial thromboplastin time, and D-dimer were measured on the STACompact Max<sup>®</sup> analyzer (Diagnostica Stago, Asnieres, France).

Thrombin–antithrombin III complex (TAT) and plasmin- $\alpha$ 2 antiplasmin complex (PAP) assays were outsourced to BioMedical Laboratories, Japan (BML Japan: <http://www.bml.co.jp>). The assay kits used were the HISCL<sup>®</sup> TAT test kit (Sysmex Corp.) for TAT assessment

**Table 1** Sequential measurement of thrombosis-related parameters after coronavirus disease-2019 vaccination

Parameters <sup>a</sup>	Baseline	Sampling day post vaccination				
		1	3	7	14	21
<i>Platelet indices</i>						
Platelet count (10 <sup>3</sup> / $\mu$ L)	168	147	159	153	168	169
PDW(fL)	8.7	7.9	8.8	8.5	9.6	8.7
MPV(fL)	8.9	8.6	8.9	8.8	9.0	9.2
P-LCR (%)	15.7	14.1	16.3	15.5	17.6	17.7
Plateletcrit (%)	0.15	0.13	0.14	0.14	0.15	0.16
TRAP aggregation (AUC unit)	101	115	104	108	104	93
<i>Coagulation</i>						
Prothrombin time (seconds)	13.3	13.6	13.4	13.3	13.0	13.8
Prothrombin time (%)	99	95	98	99	97	86
Prothrombin time (INR)	1.00	1.03	1.01	1.00	1.02	1.10
APTT (seconds)	36.0	37.1	35.8	36.3	39.1	37.5
vWF activity (%)	185	162	147	160	157	162
vWF antigen (%)	185	183	179	176	168	169
Protein C activity (IU/dL)	91	88	91	92	87	86
Protein C antigen (IU/dL)	69.2	70.4	77.5	81.5	71.6	66
Lupus anticoagulant	–	–	–	–	–	–
<i>Fibrinolysis</i>						
Plasminogen (%)	89	86	91	92	85	87
FDP ( $\mu$ g/mL)	< 0.25	< 0.25	< 0.25	< 0.25	< 0.25	< 0.25
D-dimer ( $\mu$ g/mL)	0.31	0.35	0.41	0.40	0.34	0.35

A blood sample for determining baseline values was obtained 1 day before vaccination

PDW platelet distribution width, MPV mean platelet volume, P-LCR platelet-large cell ratio, TRAP thrombin receptor-activating peptide, AUC area under the curve, INR international normalized ratio, APTT activated partial thromboplastin time, vWF von Willebrand factor, FDP fibrinogen degradation product

<sup>a</sup> Reference normal range (in parentheses): platelet count (130–450 10<sup>3</sup>/ $\mu$ L), PDW (10.0–20.0 fL), MPV (7.0–13.0 fL), P-LCR (15–35%), TRAP (92.0–151.0 AUC unit), prothrombin time (11.5–15.0 seconds), prothrombin time (70.0–135.0%), prothrombin time (0.84–1.21 INR), APTT (28.0–45.0 seconds), vWF activity (45.6–176.3%), vWF antigen (50.0–160.0%), FDP (0–5.0  $\mu$ g/mL), D-dimer (0–0.63  $\mu$ g/mL), protein C activity (72.0–160.0 IU/dL), protein C antigen (73.0–142.0 IU/dL), plasminogen (80.0–120.0 %)

and the LPIA-NV7 STACIA® test kit (LSI Medicine Corp., Tokyo, Japan) for PAP assessment.

Assays for the following parameters were outsourced to EONE Laboratories (Incheon South Korea: [https://www.eonelab.co.kr/institution/certi\\_list.asp](https://www.eonelab.co.kr/institution/certi_list.asp)): both von Willebrand factor (vWF) antigen and activity (Instrumentation Laboratory, Bedford, MA, USA); plasminogen activator inhibitor-1 (PAI-1; Asserachrom PAI-1 enzyme immunoassay kit; Diagnostica Stago); protein C and protein S antigen levels (enzyme-linked immunosorbent assay kit; Corgenix Medical Corp., Broomfield, Colorado, USA); protein C activity (synthetic chromogenic substrate method; Stachrom protein C kit; Diagnostica Stago); protein S activity (Factor Va inhibition test kit; Diagnostica Stago); lupus anticoagulant (diluted Russell viper venom test; Diagnostica Stago); fibrinogen degradation product (FDP; antibody-coated latex agglutination kit; Sekisui Medical Co. Ltd., Tokyo, Japan); and plasminogen (synthetic chromogenic substrate method; Stachrom Plasminogen kit; Diagnostica Stago).

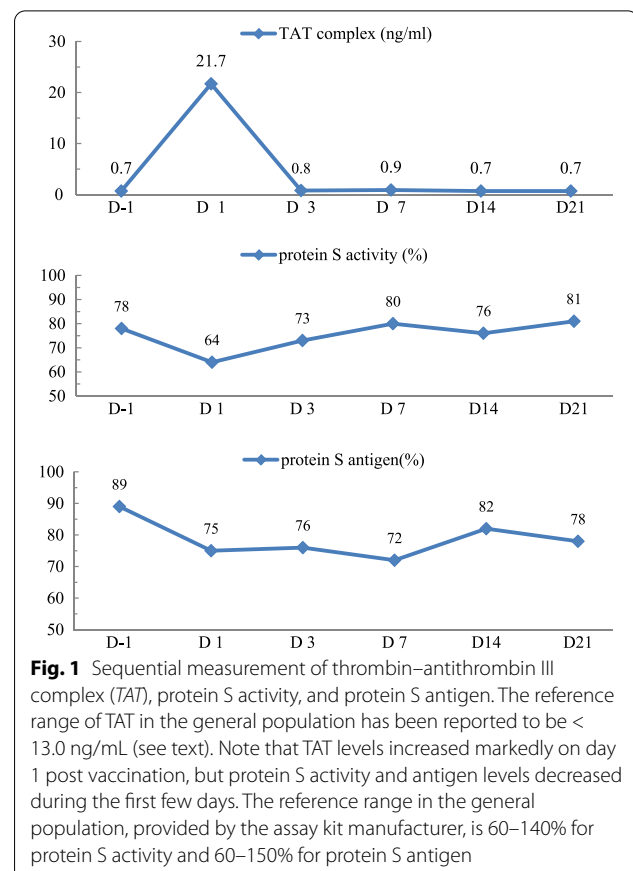
## Discussion and conclusions

It is widely acknowledged that the generation of platelet factor 4 (PF4)-polyanion antibodies is the essential pathophysiological mechanism underlying VITT development after COVID-19 vaccination [4]. However, the presence of this antibody does not always indicate thrombosis after COVID-19 vaccination [5]. What is the mechanism of PF4-polyanion antibody-negative thrombosis after COVID-19 vaccination? What kind of changes could take place in thrombosis-related parameters after COVID-19 vaccination in the general population? To answer these questions, we measured thrombosis-related parameters, before and after COVID-19 vaccination, sequentially for 21 days.

The volunteer experienced no abnormalities, except mild fever 2 days after vaccination. However, various thrombotic parameters changed during the first few days after receiving the COVID-19 vaccination (Table 1; Figs. 1, 2).

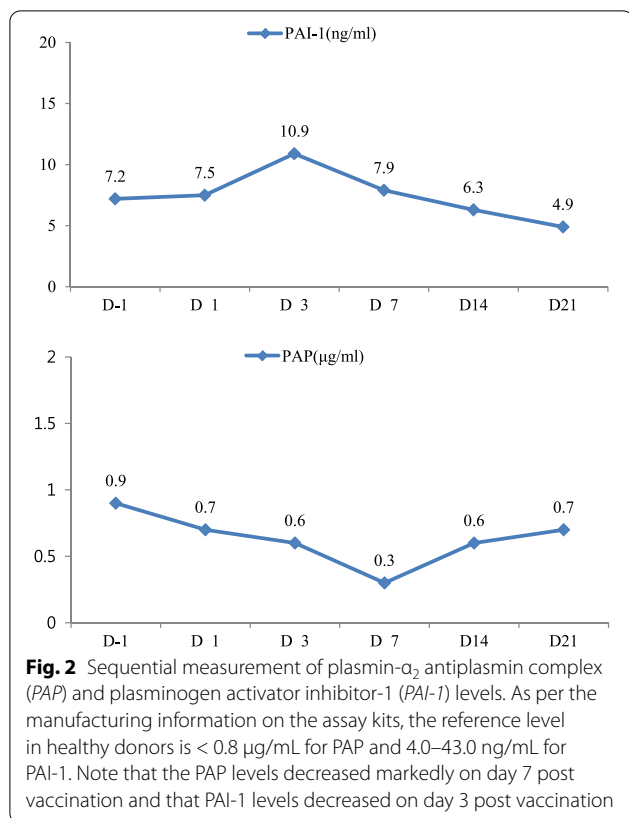
The most remarkable findings were the changes in both TAT and PAP, which indicate the generation of thrombin and plasmin, respectively [6]. The TAT level increased markedly on day 1 post vaccination from 0.7 (baseline) to 21.7 ng/mL (Fig. 1). Both the antigen level and activity of protein S decreased on days 1–3 following vaccination. As a cofactor of protein C, protein S inactivates coagulation factors Va and VIIIa. Therefore, we speculate that the AstraZeneca COVID-19 vaccine suppresses protein S, consequently provoking the generation of thrombin in circulation.

Both vWF antigen and activity decreased transiently following vaccination (Table 1; the reference ranges for



the general population, provided by the assay kit manufacturer, are 45.6–176.3% for vWF antigen and 50.0–160.0% for vWF activity). During the first few days, protein S and vWF appeared to conflict with each other in the thrombotic process, given that decreased protein S is a thrombogenic factor, while decreased vWF is an antithrombotic factor. However, increased levels of D-dimer are a surrogate marker of thrombus formation, suggesting that thrombotic activity outweighed antithrombotic activity for the first few days following COVID-19 vaccination.

There was a transient increase in PAI-1 levels from 7.2 (baseline) to 10.9 ng/mL (day 3) (Fig. 2), followed by a decrease in PAP levels from 0.9 (baseline) to 0.3 µg/mL (day 7), suggesting that plasmin generation is suppressed by PAI-1 (Fig. 2). The D-dimer level increased slightly on days 3 and 7, while the FDP level remained below the cut-off level of the assay kit (< 0.25 µg/mL) throughout the observation period. Considering the results in this case, we surmise that the fibrinolysis was not stimulated by the vaccine itself, and the reason for the increased D-dimer was a physiological response to fibrin (thrombus) formation, rather than enhanced fibrinolytic activity.



There was a transient reduction in platelet counts, from  $168 \times 10^3$  to  $147 \times 10^3/\mu\text{L}$  at 1 day after COVID-19 vaccination (Table 1). A platelet count  $< 150 \times 10^3/\mu\text{L}$  is considered as ‘thrombocytopenia,’ which is caused by either decreased production, increased destruction, or increased trapping of platelets in the spleen. We could not identify the etiology of thrombocytopenia in this case.

The volunteer was an elderly person (72 years old) with diabetes mellitus, hypertension, and hypercholesterolemia. Therefore, all the previously mentioned points, such as age and medical history, may be risk factors of thrombosis. However, we believe that if the comorbidities were the etiology underlying the change in the coagulation profile, the change would continue before and after vaccination.

As seen with platelet counts, MPV, PDW, and P-LCR were lower on post-vaccination day 1 than at baseline. Furthermore, TRAP-induced platelet aggregation increased from 101 area-under-the-curve (AUC) units to 115 AUC units after COVID-19 vaccination (Table 1), suggesting that platelet aggregation was stimulated by the COVID-19 vaccine. Further research is necessary to evaluate changes in platelet function during the first few days following COVID-19 vaccination.

This study has its limitations. This report only describes the case of a single volunteer, and we only measured selective thrombotic and antithrombotic parameters. Therefore, it is uncertain whether our observations can be applied to other cases. However, even with this limitation, our observation indicates that various thrombotic parameters may change during the first few days after COVID-19 vaccination, without signs of clinically overt thrombosis.

Increased thrombotic factors (such as decreased protein S) with decreased fibrinolytic activity due to increased PAI-1 seem to be potential factors causing thrombogenesis after COVID-19 vaccination. To better understand the progression of thrombosis, sequential measurement of PI, TAT, PAP, protein C, protein S, vWF, D-dimer, and PAI-1 after COVID-19 vaccination seems to be informative.

#### Abbreviations

COVID-19: Coronavirus disease 2019; FDP: Fibrinogen degradation product; MPV: Mean platelet volume; PAI-1: Plasminogen activator inhibitor-1; PAP: Plasmin- $\alpha_2$  antiplasmin complex; PDW: Platelet distribution width; PF4: Platelet factor 4; PI: Platelet indices; P-LCR: Platelet-large cell ratio; TAT: Thrombin-antithrombin III complex; TRAP: Thrombin receptor-activating peptide; VITT: Vaccine-associated immune thrombosis and thrombocytopenia; vWF: Von Willebrand factor.

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#### Author contributions

Conceptualization: SJ, SS, SH. Data curation: JR. Original draft, reviews and edits: SH. Supervision: JL. All authors read and approved the final manuscript.

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#### Availability of data and materials

Not applicable.

#### Declarations

#### Ethics approval and consent to participate

The case report protocol was reviewed and approved by the Public Institutional Review Board of the Ministry of Health and Welfare of South Korea (<http://irb.or.kr/menu02/summary.aspx>, approval no.: P01-202107-21-018).

#### Consent for publication

Written informed consent was obtained from the patient for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

#### Competing interests

The authors declare that they have no competing interests.

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## References

1. Greinacher A, Thiele T, Warkentin TE, Weisser K, Kyrle PA, Eichinger S. Thrombotic thrombocytopenia after ChAdOx1 nCov-19 vaccination. *N Engl J Med*. 2021;384:2092–101. <https://doi.org/10.1056/NEJMoa2104840>.
2. World Health Organization. Coronavirus disease (COVID-19) pandemic. <https://www.who.int/emergencies/diseases/novel-coronavirus-2019?gclid=CjwKCAjwoZWH>. Accessed 14 Dec 2021.
3. Levi M, van der Poll T. Coagulation and sepsis. *Thromb Res*. 2017;149:38–44. <https://doi.org/10.1016/j.thromres.2016.11.007>.
4. Thiele T, Ulm L, Holtfreter S, et al. Frequency of positive anti-PF4/polyanion antibody tests after COVID-19 vaccination with ChAdOx1 nCoV-19 and BNT162b2. *Blood*. 2021;138:299–303. <https://doi.org/10.1182/blood.2021012217>.
5. Favaloro EJ, Henry BM, Lippi G. The complicated relationships of heparin-induced thrombocytopenia and platelet factor 4 antibodies with COVID-19. *Int J Lab Hematol*. 2021;43:547–58. <https://doi.org/10.1111/ijlh.13582>.
6. Lundbeck M, Krag AE, Christensen TD, Hvas AM. Thrombin generation, thrombin-antithrombin complex, and prothrombin fragment F1+2 as biomarkers for hypercoagulability in cancer patients. *Thromb Res*. 2020;186:80–5. <https://doi.org/10.1016/j.thromres.2019.12.018>.

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